

Detection virulence factors of *Klebsiella pneumoniae* from cattle by using PCR tec.

Jameela Radi Esmaeel

Jinan Nadhim Sadeq

Coll. of Vet. Med. / Univ. of Al-Qadisiyah

email: Jameela.Esmaeel@qu.edu.iq

Abstract

Identification of *K. pneumoniae* was evaluated using conventional microbiological characteristics and molecular assays .

Milk specimens were collected from cattle suffered from clinical symptoms of mastitis from different farms of AL-Qadisiyah province.

Out of 45 samples obtained, 20 isolates (44.4 %) were detected as *K. pneumoniae* according to morphology of colonies and biochemical features.

Molecular detection of *Klebsiella pneumoniae* based 16S rRNA gene for determination two virulence factor genes(*rmpA*,*magA*) by using specific primer.These genes potent the pathogenesis of *Klebsiella pneumoniae*. The primers were made in this study by using NCBI-GenBank and primer3 plus design online. The primer is made by company in Korea (Bioneer).

Molecular detection of isolates which give away specific PCR products of of 312bp for *magA* gene and 835bp for *rmpA* gene .

The *magA* and *rmpA* genes were amplified in six (30%)and five(25%) isolates out of 20 isolates of *Klebsiella pneumoniae*.

Keywords: cattle mastitis, Virulence factors, *Klebsiella. pneumoniae*,PCR tech.

الكشف عن عوامل ضراوة الكلبسيلا الرئوية المعزولة من الابقار باستخدام تقنية سلسلة تفاعل البلمرة

جنان ناظم صادق

جميلة راضي اسماعيل

كلية الطب البيطري/ جامعة القادسية

الخلاصة

تم تشخيص الكلبسيلا الرئوية الخصائص الميكروبيولوجية التقليدية وطرق القياس الجزيئي . تم جمع عينات الحليب من ابقار تعاني من علامات سريرية لالتهاب الضرع من حقول مختلفة في محافظة القادسية . من بين 45 عينة تم الحصول عليها .تم الكشف عن 20 عزلة بنسبة (44,4%). وتم تحديد الكلبسيلا الرئوية على اساس شكل المستعمرات والاختبارات الكيموحيوية .

تم التشخيص الجزيئي للكلبسيلا الرئوية 16srRNA بالاستناد على جين باستخدام بادئات متخصصة (MagA,rmpA) لتحديد عاملين من عوامل الضراوة وهما صممت هذه البادئات في الدراسة باستخدام بنك الجينات وصممت ثلاث بادئات جهزت بواسطة كوربا .تم الكشف الجزيئي للعزلات وحددت نواتج

(Bioneer company) شركة

312 و 835) للجينين على التوالي . PCR bp

ظهر تضخيم الجينات في ستة عزلات بنسبة (30%) وخمسة عزلات بنسبة (25%) من اصل 20 عزلة من الكلبسيلا الرئوية .

Introduction

Small number of bacterial species is responsible for most mastitis cases classified into contagious and environmental(1).

Klebsiella pneumonia is present in the normal animals , its an important cause of acquired infection and its one of major organisms among of the gram negative that cause mastitis specially in cattle(2,3) .

Mastitis caused by *K. pneumoniae* is poor response to antibiotic therapy so that case of clinical mastitis caused by it is more sever (4).

Klebsiella pneumonia can arise from cows in environment enter to the udder through milking ,or teats when contamination by feces ,mud and bedding materials(5).

The presence of virulence genes in *Klebsiella pneumonia* promote the pathogenesis to evading the immune of the body (6) .

Many sequenced virulence genes have been detection in *Klebsiella pneumonia* such as 16 sRNA ,Mag A and rmpA. analysis and Sequencing of regions within 16S rRNA gene can expand speedy and effective ways to estimate variety of bacteria useful for pathogen and identification(7)

Mag A(mucoviscosity-associated gene A)is causes mucoviscosity,also is encodes apslar polymerase and find within gene specific *Klebsiella pneumonia* (8) .

While the rmpA gene is related with hypermucoviscous phenotype and encods as plasmid borne regulator of extracellular polysaccharide synthesis, a layer arounding the surface of cell, is hinders the phagocytosis (9)

these two genes associated with invasive infections (10) .

The aim of this study is to determine some virulence genes of *Klebsiella pneumonia* isolated from clinical case of mastitis in cattle in AL-Qadisyiah province via microbiological characteristics and molecular methods .

Materials and Methods

This study was carried out in the veterinary medicine collage\ university of AL-Qadisyiah .

Sample collection:The specimens were milk, collected from cattle undergoing suffers from clinical sings of mastitis from different areas of AL-Qadisyiah province.

All milk samples were cultured on maCconkeys agar and blood agar blates and incubated for 24 hours at 37c according to standerd procedure (11) .

Thereafter the isolates were activated by in inoculated on CHROMagar Orintation and incubated at 37C° for overnight.

Identification of isolates based on morphology of colonies ,subculturing of isolates onto maCconkey and incubated for 24 hours at 37c, pink ,mucoid,lactose fermented colonies were considered to be *Klebsiella* spp,On orintation media colonies is metallic blue color,large ,rounded .

The identification of species of *Klebsiella* according to the biochemical reaction such as catalase,oxidase, indole production ,motility and citrate utilization . biochemical reactions carried out according to(12) .

Bacterial genomic DNA extraction:

Bacterial genomic DNA was extracted from *Klebsiella pneumoniae* isolates by

using (Presto™ Mini gDNA Bacteria Kit. Geneaid. USA). 1ml of overnight bacterial growth on BHI broth was placed in 1.5ml microcentrifuge tubes and then transferred in centrifuge at 10000 rpm for 1 minute. After that, the supernatant was discarded and the bacterial cells pellets were used in genomic DNA extraction and the extraction was done according to company instruction. After that, the extracted gDNA was checked by Nanodrop spectrophotometer, then store

in -20C at refrigerator until perform PCR assay.

Polymerase chain reaction (PCR):

PCR assay was performed for confirmative detection of *Klebsiella pneumoniae* based 16S rRNA gene and for determination some virulence factor genes and by using specific primer that designed in this study by using NCBI-GenBank and primer3 plus design online. The primers were provided by (Bioneer company . Korea) as table(1).

Primers	Sequences	Amplicon	GenBank
16S rRNA	CGCGAAGAACCTTACCTGG T	352bp	Y17669.1
	AGTTGCAGACTCCAATCCG G		
MagA	TAGGTCAGGCAGCTGTTGT G	312bp	KP973856.1
	GCTCCGTTGCAATATGACC G		
RmpA	TGCAAACACGCAAAGGACA A	835bp	AB289644.1
	AAGAGTGCTTTCACCCCT C		

Table (1): sequences of primers with size of targeted products of the PCR to the *K. pneumoniae*

Prepared of the PCR master mix by using (kit of AccuPower® PCR PreMix). from company Bioneer in Korea. Premix tubes of the PCR contain freezer-dried pellet to (dNTPs 250µM ,Tris-HCl (pH 9.0),Taq DNA polymerase 1U, 10mM, KCl 30mM, stabilizer ,MgCl₂ , tracking dye , and1.5mM) prepared of the PCR master mix according to the kit of instructions with 20µl total volume ,added 5µl of genomic DNAwas purified and also added 1.5µl of 10p mole of forward primer with same of reverse primer, The PCR premix tube by PCR water (20µl) mixed by Exispin vortex centrifuge. The reaction was carried out in a thermocycler by set up the following thermocycler conditions; initial denaturation temperature of 95 C° for 5 min; followed by 30 cycles at denaturation 95 C° for 30 s, annealing 58 C° for 30 s, and extension 72 °C for 1min and then final extension

at 72 °C for 10 min. The PCR products were examined by electrophoresis in a 1% agarose gel.

Results

growth on Orientation medium where specific color give blue pigment, The growth was also inoculated onto MacConkey agar, where lactose fermenting mucoid colonies producing pink of the medium lactose fermenting. These isolates were positive citrate utilization and negative for H₂S production positive for catalase and negative for oxidase reaction according to (11)

In the present study, the prevalence of *Klebsiella pneumonia* was found to be 44.4%.



Figure 1: *Klebsiella pneumonia* on MacConkey agar produced rounded, mucoid (polysaccharide capsule), large and pink colonies

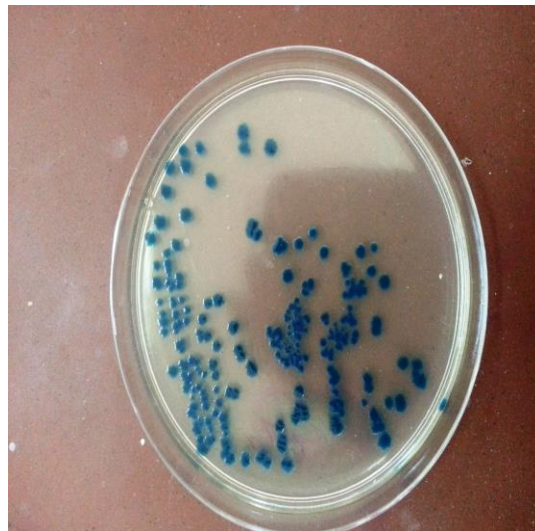


Figure 2: *Klebsiella pneumonia* on chrom agar Orientation, produce metallic blue, rounded and large colonies

In this study, *magA* gene, were tested by PCR by using specific primer sequences with product sizes of 312bp ,Out of total 20 isolates, 6 isolate (30%) was positive for *magA* gene .

While gene *rmpA* with product sizes of 835bp, Out of total 20 isolates, clarify that 5 isolate(25%) was positive for *rmpA* gene while 15 isolates of *Klebsiella* (75 %) were negative to this gene.

Gene	No .of tested isolates	Positive
16S rRNA gene	20	20
MagA	20	6
RmpA	20	5

Table (2):The virulence genes distribution with the numbers of the isolates

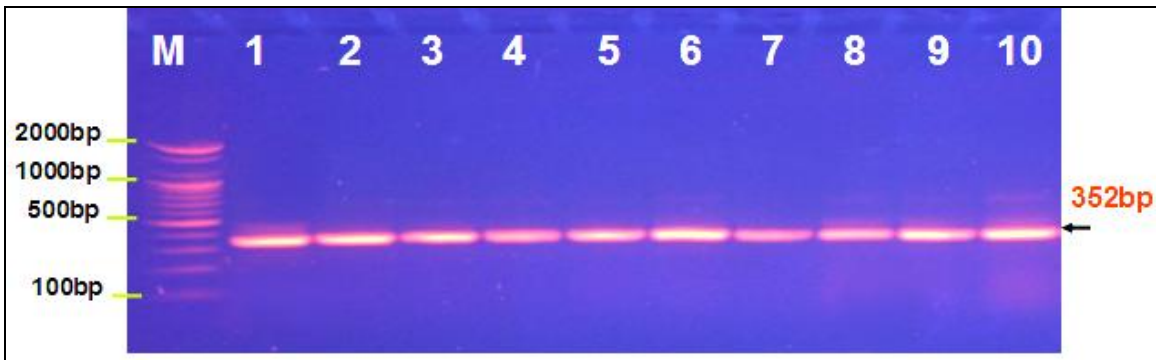


Figure 3: Agarose gel electrophoresis image that show the PCR product analysis of 16S rRNA gene in *Klebsiella pneumoniae* positive isolates. Where M: marker (2000-100bp), lane (1-10) positive *Staphylococcus aureus* at (352bp) 16S rRNA gene PCR product.

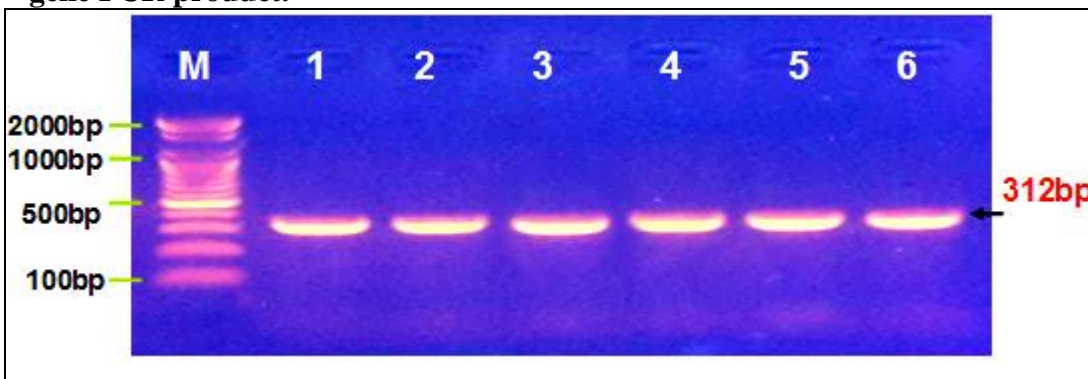


Figure 4: Agarose gel electrophoresis image that show the PCR product analysis of *magA* gene in *Klebsiella pneumoniae* isolates. Where M: marker (2000-100bp), lane (1-6) positive isolates at (312bp) PCR product.

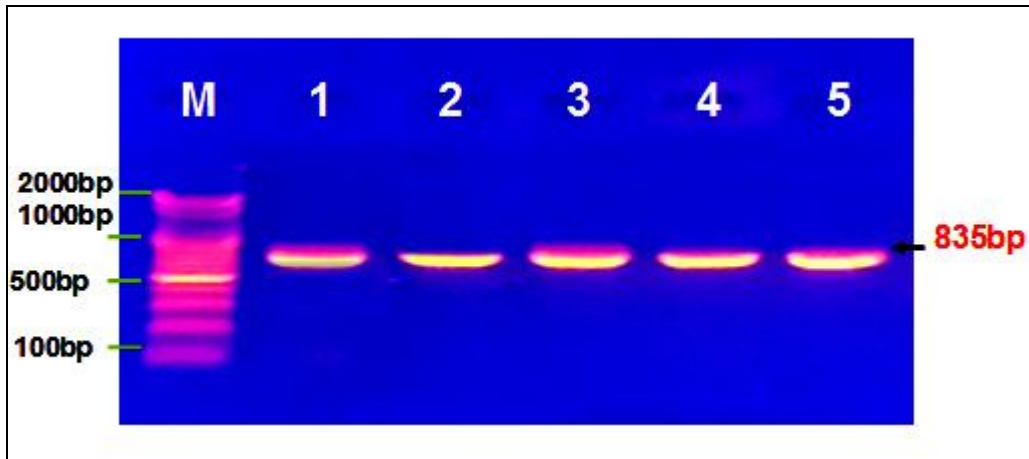


Figure 5: Agarose gel electrophoresis image that show the PCR product analysis of *rmpA* gene in *Klebsiella pneumoniae* isolates. Where M: marker (2000-100bp), lane (1-5) positive isolates at (835bp) PCR product.

Discussion

The most frequent problem in the farms of dairy cattle in the world is mastitis (13). The occurrence of mastitis in cattle differ according to the regions due to religious law, also varies according to management accounting, and hygiene measures.

In clinical mastitis the most common gram negative bacteria is *Klebsiella pneumoniae* (14).

shedding of *Klebsiella* spp in feces promote the distribution of pathogens in the environment, material of bedding and then suggested recycled transmission of *Klebsiella pneumoniae* in dairy farms. (15).

(16) found that large number of mastitis outbreaks by *Klebsiella pneumoniae*.

The contamination of milk may result from contact with cow which infected or may be

contact with contamination of environment (17).

The occurrence of mastitis by *Klebsiella* increases under humid and hot conditions (18)

outbreaks by *Klebsiella* are reported in many countries in highly percent exemplify USA and Eroupe (19).

sample collection methods may play a role in heterogeneity of *Klebsiella* in milk samples

Our high rates are proximate to the results in south America and Asia which have

registrated in rates between 33.5% and 45% (20)(21), While the findings of other study in

france a high values which have registrated 92%. (22). The pathogenic strains of

Klebsiella pneumoniae existent in the environment in broad variety (23).

(24) were observed that both of serotypes of capsular related to *Klebsiella pneumoniae*

which cause mastitis and human consider as source of infection.

Species of *klebsiella* can be detected by 16SrRNA sequencing of gene, but due to the nucleotide variation is limited, 16SrRNA sequences not be used to differentiation within the phylogenetic groups (22), So 16S rDNA analysis of

phylogenetic used in classifying and identifying *K. pneumoniae*.

MagA and *rmpA* causing the bacteria is the most invasive and resistant to immune defence (25)

A prior studies has shown that the phenotype of mucoid colony may be to a gene

specified *rmpA* (26).

in our study gene *rmpA* was detected in only 5 isolates, this less from previous study (27)

The public health importance of this study is to know unique regulator of hypermucoviscous capsule composition and formation (28).

Regarding analysis of molecular of *Klebsiella pneumoniae* confirmed in highly specificity of primers from region of 16SrRNA.

Conclusion

((1)) molecular methods measurement instrument helpful in diagnostics of mastitis ((2)) control of *Klebsiella* in farms of dairy cattle by hygiene ((3)) Detection of the virulence factors in *K. pneumoniae* that cause mastitis in cattle will be aid in detection of this disease .

References

- 1- Langoni H.2013 Qualidade do leite: utopia sem um programa sério de monitoramento da ocorrência de mastite bovina. *Pesq. Vet. Bras.* 33(5): 620-626.
- 2- Locatelli C, Scaccabarozzi L, Pisoni G, Moroni P. CTX-M1 ESBL-producing *Klebsiella pneumoniae* subsp. *pneumoniae* isolated from cases of bovine mastitis. *J Clin Microbiol.* 2010;48:3822-3.
- 2- Locatelli C, Scaccabarozzi L, Pisoni G, Moroni P. CTX-M1 ESBL-producing *Klebsiella pneumoniae* subsp. *pneumoniae* isolated from cases of bovine mastitis. *J Clin Microbiol.* 2010;48:3822-3.
- 3-Schukken Y, Chuff M, Moroni P, Gurjar A, Santisteban C, Welcome F, et al. The 'other' Gram-negative bacteria in mastitis: *Klebsiella, serratia*, and more. *Vet Clin North Am: Food Anim Prac.* 2012;28:239-56.
- 4- Silva N. & Costa G.M. 2001. An outbreak of acute bovine mastitis caused by *Klebsiella pneumoniae* in a dairy herd.. *Arq. Bras. Med. Vet. Zootec.* 53(4):1-5.
- 5-FAOSTAT-Agriculture (food and Agriculture organization statistics)Annual Agricultural data .2007., Accessed may 20,2011.
- 6-Elliott S J, Wainwright L A, McDaniel T K, Jarvis K G, Deng Y K, Lai L C, McNamara B P, Donnenberg M S, Kaper J B. The complete sequence of the locus of enterocyte effacement (LEE) from enteropathogenic *Escherichia coli* E2348/69. *Mol Microbiol.*1998;28:1-4
- 7- Kolbert CP, Persing DH (1999) Ribosomal DNA sequencing as a tool for identification of bacterial pathogens. *Curr Opin Microbiol* 2(3): 299-305
- 8-Yeh KM, Lin JC, Yin FY, Fung CP, Hung HC, Siu LK, Chang FY. 2010. Revisiting the importance of virulence determinant *magA* and its surrounding genes in *Klebsiella pneumoniae* causing pyogenic liver abscesses: exact role in serotype K1 capsule formation. *J. Infect. Dis.* 201:1259-1267. .10.1086/606010 .
- 9-Doyle J and Evans. Jr. (2008)"*Klebsiella*". *J AccessScience.*33: 100-300.
- 10-Yu WL, Ko WC, Cheng KC, Lee HC, Ke DS, Lee CC, Fung CP, Chuang YC. 2006. Association between *rmpA* and *magA* genes and clinical syndromes caused by *Klebsiella pneumoniae* in Taiwan. *Clin. Infect. Dis.* 42:1351-1358. .10.1086/503420 .

- 11-Quinn P.J., Markey B., Carter M.E., Donnelly W.J. & Leonard F.C. 2005. *Microbiologia Veterinária e Doenças Infecciosas*. Artmed, Porto Alegre
- 12- MacFaddin. (2000). *Biochemical tests for identification of medical bacteria*. 3rd edition". The Williams and Wilkins-Baltimore, USA. Philadelphia.
- 13-Tiwari JG, Babra C, Tiwari HK, Williams V, de Wet S, Gibson J, et al. Trends in therapeutic and prevention strategies for management of bovine mastitis: an overview. *J Vacc Vaccinol*. 2013;4:176
- 14- Munoz M.A. & Zadoks R.N. 2007. Patterns of fecal shedding of *Klebsiella* by dairy cows. *J. Dairy Sci.* 90:1220-1224.
- 15- Schukken Y, Chuff M, Moroni P, Gurjar A, Santisteban C, Welcome F, et al. The 'other' Gram-negative bacteria in mastitis: *Klebsiella*, *serratia*, and more. *Vet Clin North Am: Food Anim Prac*. 2012;28:239–56.
- 16-Langoni H., Corrêa C.N.M., Corrêa W.M., Barros J.A. & Corrêa G.N. 1985. Mastites bovinas por *Candida* e *Klebsiella* *Revta Bras. Med. Vet.* 7(7): 203-204.
- 17-Jasper, D. E., and J. D. Dellinger. 1975. Teat apex coliform populations and coliform mastitis—a herd study. *Cornell Vet.* 65:380-392.
- 18-Hogan, J., and K. L. Smith. 2003. *Coliform mastitis*. *Vet. Res.* 34:507-519.
- 19- Paulin-Curlee GG, Singer RS, Sreevatsan S, Isaacson R, Reneau J, Foster D, et al. Genetic diversity of mastitis-associated *Klebsiella pneumoniae* in dairy cows. *J Dairy Sci.* 2007;90:3681–9.)(Vecht U, Meijers KC, Wisselink HJ. *Klebsiella pneumonia* mastitis as a dairying problem. *Tijdschr Diergeneeskd.* 1987;112:653–9.
- 20- Pitkala A, Haveri M, Pyorala S, Myllys V, Honkanen-Buzalski T. Bovine mastitis in Finland 2001 — prevalence, distribution of bacteria, and antimicrobial resistance. *J Dairy Sci.* 2004;87:2433–41.
- 21- Botrel MA, Haenni M, Morignat E, Sulpice P, Madec JY, Calavas D. Distribution and antimicrobial resistance of clinical and subclinical mastitis pathogens in dairy cows in Rhône-Alpes, France. *Foodborne Pathog Dis.* 2010;7:479–87.
- 22-Brisse, S., and J. Verhoef. 2001. Phylogenetic diversity of *Klebsiella pneumoniae* and *Klebsiella oxytoca* clinical isolates revealed by randomly amplified polymorphic DNA, *gyrA* and *parC* genes sequencing and automated ribotyping. *Int. J. Syst. Evol. Microbiol.* 51(3):915–924
- 23- Munoz MA, Welcome FL, Schukken YH, Zadoks RN. Molecular epidemiology of two *Klebsiella pneumoniae* mastitis outbreaks on a dairy farm in New York State. *J Clin Microbiol.* 2007;45:3964–71.
- 24- Johnson JG. Regulation of Type 3 fimbrial gene expression in *Klebsiella pneumoniae* [dissertation] Iowa City, IA: The University of Iowa; 2011.
- 25- Shon AS, Bajwa RP, Russ TA. Hypervirulent (hypermucoviscous) *Klebsiella pneumoniae*: a new and dangerous breed. *Virulence.* 2013;4:107–18.
- 26- Nassif X, Honore N, Vasselon T, Cole ST, Sansonetti PJ. Positive control of colanic acid synthesis in *Escherichia coli* by *rmpA* and *rmpB* two virulence-plasmid genes of *Klebsiella pneumoniae*. *Mol Microbiol.* 1989;3:1349–59.

27- Brisse S, Fevre C, Passet V, Issenhuth-Jeanjean S, Tournebize R, Diancourt L, Grimont P. 2009. Virulent clones of *Klebsiella pneumoniae*: identification and evolutionary scenario based on genomic and phenotypic characterization. PLoS One 4:e4982. [.10.1371/journal.pone.0004982](https://doi.org/10.1371/journal.pone.0004982).

28-Valley,(2011) *rmpA* a Unique Virulence Regulator in Emerging *Klebsiella pneumoniae*. J. Infectious Diseases. p 10-069.