Gross and histological comparison of hydatid cyst infection in livers of sheep and cows

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Abstract

3 samples of fertile hydatid cysts and 3 other sterile were collected from sheep liver and 3 samples of sterile hydatid cysts from cows liver. Infected liver samples with hydatid cyst were obtained from meat seal shops in AL-Muthanna province in December 2010, then measured dimensions of cysts and calculated quantity of hydatid fluid and viability of protoscolices by staining with eosin stain then examining under light microscope of 40x. petechial hemorrhage and paleness around cysts were important gross lesions in sheep in addition to calcification in cow hydatid cysts, on other hand the histological lesions were thickened fibrous layer and sever necrosis with infiltrations of eosinophils and extensive calcification in cows liver. There was significant effect of amount of hydatid cyst, thickness of fibrous layer and thickness of necrotic layer.

Introduction

Hydatid disease due to cystic echinococcosis is one of the most important public health and economic problems in different countries including Iraq. Hydatid disease is a zoonosis caused by the tapeworm of Echinococcus spp. These species include E. granulosus, E multilocularis, E vogeli or E oligarthrus. These parasites live as a small intestinal tapeworm of dogs and occasionally other carnivores. Their larval form causes hydatidosis in domestic animals and man. The shedding of gravid proglottids or eggs in the feces occurs within 4–6 weeks after infection of the definitive host. Ingestion of eggs by intermediate host animals (sheep, cattle, goats, horses, camels) or human results in the release of an oncosphere into the gastrointestinal tract, which then migrates to primary target organs such as liver and lungs, and less frequently to other organs. Usually the fully mature metacestode (i.e. hydatid cyst) develops within several months or years. The hydatid disease is prevalence where livestock is raised in association with dogs. Those endemic areas include Australia, Latin America, Europe, Africa and the Middle East. Liver is the most common site of cystic development, in over 90% of liver cysts; the oncosphere is trapped in the central veins of the hepatic lobules and the resultant cyst may be deep or superficial and it causes compression of the liver cells which can lead to biliary stasis and cholangitis. The cyst may present as a liver abscess and large cyst can produce localized or diffuse hepatomegaly. Local pathological effects depend on the site of the hydatid cyst; ruptured liver cyst through the diaphragm can produce a pleural effusion or bronchobiliary fistula. The parasite destroys the liver parenchyma, bile ducts and blood vessels resulting in symptoms of biliary obstruction, portal hypertension and necrosis of the central portion of the cyst with abscess formation. Growth of the germinal membrane into blood vessels produces metastasis to almost any organ, but they are more commonly found in the lungs and brain.

Materials and Methods

3 samples of fertile hydatid cysts and 3 other sterile were collected from sheep liver and 3 samples of sterile hydatid cysts from cow’s liver. Infected liver samples with hydatid cysts are obtained from meat seal shops in AL-Muthanna province in December 2010 and put the samples in cleaned container and transferred to laboratory. In first time important gross pathological changes recorded on infected samples like petechial hemorrhage, infiltrations, bloody patches around cysts
and calcifications. Then measured (length and width) of cysts and calculated the quantity of hydatid fluid and collected in clean and disinfected containers. Hydatid fluid was aspirated from cyst by syringe and protoscolices were scraped from sides of germinal layer and putting in test tubes then centrifuged at 2500 rpm for 5 minutes, the supernatant discharged and sediment used for measuring viability of protoscolices by staining with eosin stain then examining under light microscope of 40x. the red protoscolices consider dead while other green are alive. Cysts opened carefully and noticed germinal layer which isolated in container then cutting the cyst with part of liver tissue for certain distances (1cm, 2cm and 3cm) from cysts and tissues put in formalin 10% then discharging of formalin and replaced with another formalin after one day. SAS Users Guide used with Complete Randomized Design in analyzing of research data, the differences between averages were tested at P≤0.05 by Duncan.

**Results**

The samples were evaluated grossly and histology.

**Grossly**

**Sheep**

Sterile cysts: petechial hemorrhage and infiltrations inflammatory cells around cysts.

Fertile cysts: blood patches, infiltrations of inflammatory cells around cysts and paleness of most areas of liver.

**Cows**

Sterile cysts: calcifications, blood patches and infiltrations of inflammatory cells around cysts and pale on margins of liver. Length, width and amount of fluid in sheep for sterile hydatid cysts were 5.25 centimetres (cm), 2.50 cm and 16.33 millilitres (ml) respectively while for fertile hydatid cysts were 5.50 cm, 3.50 cm and 27.80 ml respectively. In cows for sterile hydatid cysts were 5.50 cm, 2.50 cm and 8.58 ml respectively (table 1,2) (figure 1,2). The significant effect exist at p≤0.05 of amount of fluid of hydatid cysts.

**Histology**

**Sheep:**

Sterile cysts: thickened fibrous layer and sever necrosis with eosinophilia and severe scattered of inflammatory cells with fibrous tissue. After these layers, the normal tissue appears with foci of inflammatory cells (picture 7,8).

**Cows**

Sterile cysts: Thickened fibrous layer and sever necrosis with eosinophilia and extensive calcification along hepatic tissue. The normal tissue appears with severe scattered of inflammatory cells and fibrous tissue (picture 9,10). Thickness of fibrous layer of sterile and fertile hydatid cysts in sheep and sterile cysts in cows were 47.50 micrometer, 48.42 micrometer and 93.94 micrometer respectively, thickness of necrotic layer in sheep of sterile and fertile hydatid cysts and sterile cysts in cows were 61.15 micrometer, 73.17 micrometer and 45.50 micrometer respectively. The significant effect exist at p≤0.05 (table 3) (figure 3).

| Table (1) length of sterile and fertile hydatid cysts in sheep and cows |
|-------------------------------------------------|---------------------------------|-----------------|-----------------|-----------------|
| Sheep                                          | cow                            | sterile cysts   | fertile cysts   | N.S.            |
| length of cysts(cm)                            | 5.25                           | 5.50            | 5.50            |                 |
| width of cysts(cm)                             | 2.50                           | 3.50            | 2.50            |                 |

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Table (2) amount of fluid of sterile and fertile hydatid cysts in sheep and cows

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<td>fertile cysts</td>
<td>sterile cysts</td>
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<td>amount of fluid of cysts (cm$^3$)</td>
<td>b*</td>
<td>a*</td>
<td>c*</td>
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<td>16.33</td>
<td>27.80</td>
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*The different letters in the same row refer to presence of significant effect at $p \leq 0.05$.

Fig. (1) length and width of sterile and fertile hydatid cysts in sheep and cows

Fig. (2) amount of fluid (cm$^3$) of sterile and fertile hydatid cysts in sheep and cows
Fig. (3) thickness of fibrous layer and necrotic layer (micrometer) of sterile and fertile hydatid cysts in sheep and cows

Pic. (1) Liver of sheep infected with hydatid cyst
Pic. (2) Brood capsule from infected sheep liver by eosin (Red)
Pic. (3) Dead protoscoleces stained by eosin (Red)
Pic. (4) Alive protoscolices stained by eosin (Green)
Pic. (5) thickened fibrous layer (f) and severe necrosis (n) of sterile cysts in sheep

Pic. (6) thickened fibrous layer (f) and severe necrosis (n) of sterile cysts in sheep

Pic. (7) fibrous layer (f) and severe necrosis with eosinophilia (n) of fertile cyst in sheep

Pic. (8) fibrous layer with severe scattered of inflammatory cells with fibrous tissue (f) of fertile cyst in sheep
Discussion

The present study revealed that the presence of hydatid cysts in the liver of sheep and cows resulted in different histological lesions represented in inflammatory reaction, fibrosis and necrosis in the area near to the cysts. Such of these effects were mostly documented in different animals infected with different species of *Echinococcus* [6,14,15]. In goat and sheep, Blanton et al. (1998) [16] showed evidence of marked host cellular reaction consisting of infiltration of the adventitial layer with neutrophils, eosinophils, and plasma cells. In addition to this inflammatory infiltrate, the new space between the liver tissue and cyst wall contained disorganized fibroblasts and mesenchymal cells. In the most necrotic areas, the laminate layer could not be collected together with adherent liver tissue and the adventitial layer appeared completely degenerated and it was replaced by acute inflammatory cells. Also, Ritter (1987) [17] suggested that the liver cell necrosis may be either due to progressive action of intracellular enzymes of the injured cells or to a metabolic disturbance and inhibition of synthesis needed of DNA and hence protein synthesis. There is significant effect of amount of hydatid cyst which is higher in fertile cyst that contain alive and proliferated protoscolices which based on hydatid fluid components in nutritional requirements. The thickness of
fibrous and necrotic layer is significantly affect between sheep and cows due to different of defense mechanism and cellular effects with difference of animal species. In conclusion, echinococcosis is still an important health problem in Iraq that needs further study. The people traveling to endemic areas should be educated not to contact with wild animals. There is a need for a suitable eradication program, so that untreated dogs as well as foxes, which play an important role in the contagiousness of the disease in rural areas, can be eradicated as an infectious agent.

References

مقارنة عيانية ونسبيّة للإصابة بالأكياس العذرية في أكباد الأغنام والأبقار

منصور جدعان علي الخالدي
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الخلاصة
تُأخذ عينتين من أكياس عذرية من أكباد أغنام (3 عينات عميقة ومثلها خصبة) و3 عينات من الأكياس العميقة في أكباد الأبقار. وُضعت في محلات القمرين في محافظة المثنى. كان القمرين الأول لسنة 2010. فُست أبعاد الأكياس وحسبت كمية وحية الرؤوس الأولية باستخدام صبغة الأبيسين تحت المجهر الضوئي بالقوة X40. ثم أخذت مقاطع نسيجية لهذه الأكياس ومساحات معينة وقُرب التأثير المحلي لهذه الأكياس على أنبوبة الكبد وأكياس الصفرا. وُجد أن النزف الحبري والشحوب في أكباد الأغنام المصدر بالإضافة للتلكس في أكباد الأبقار أهم الأتفاق العنيفة المشاهدة. النتاج النسيجي فقد لوحظت للكوينية السميكة وطقة النخر. وارتشاح الخلايا الحمضية والتكليس الواسع الشامل لكل أجزاء المنطقة المصابة في أكباد الأبقار المصابة. ظهير التأثير المعنوي في كمية السائل العدي وسمكة الضيقة السميكة وسمكة طبقة النخر.